

# Development of organ on chip technology for predictive toxicology and applied pharmacology



**Eric Leclerc**

**CNRS**

**The University of Tokyo**

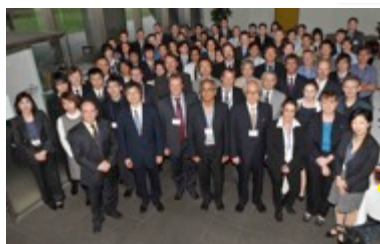
**Universite de Technologie de Compiegne**



IIS in old Roppongi Campus



1995



2001

Komaba II Campus

International Unit  
UMI 2820

2004

2008

2011: The Great  
Earthquake

2012

Mirror structure  
in Hospital

2016

2014

iLite RHU program

New research axis

2016



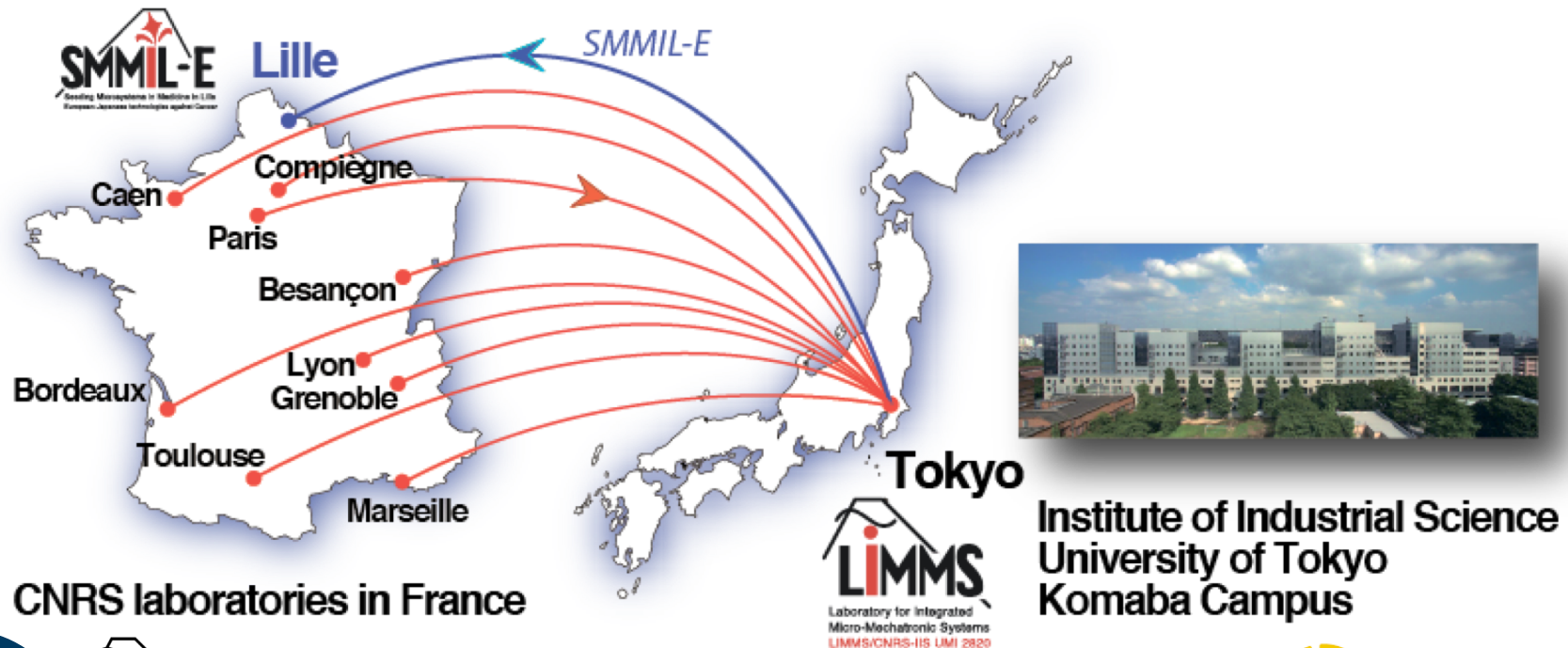
Today



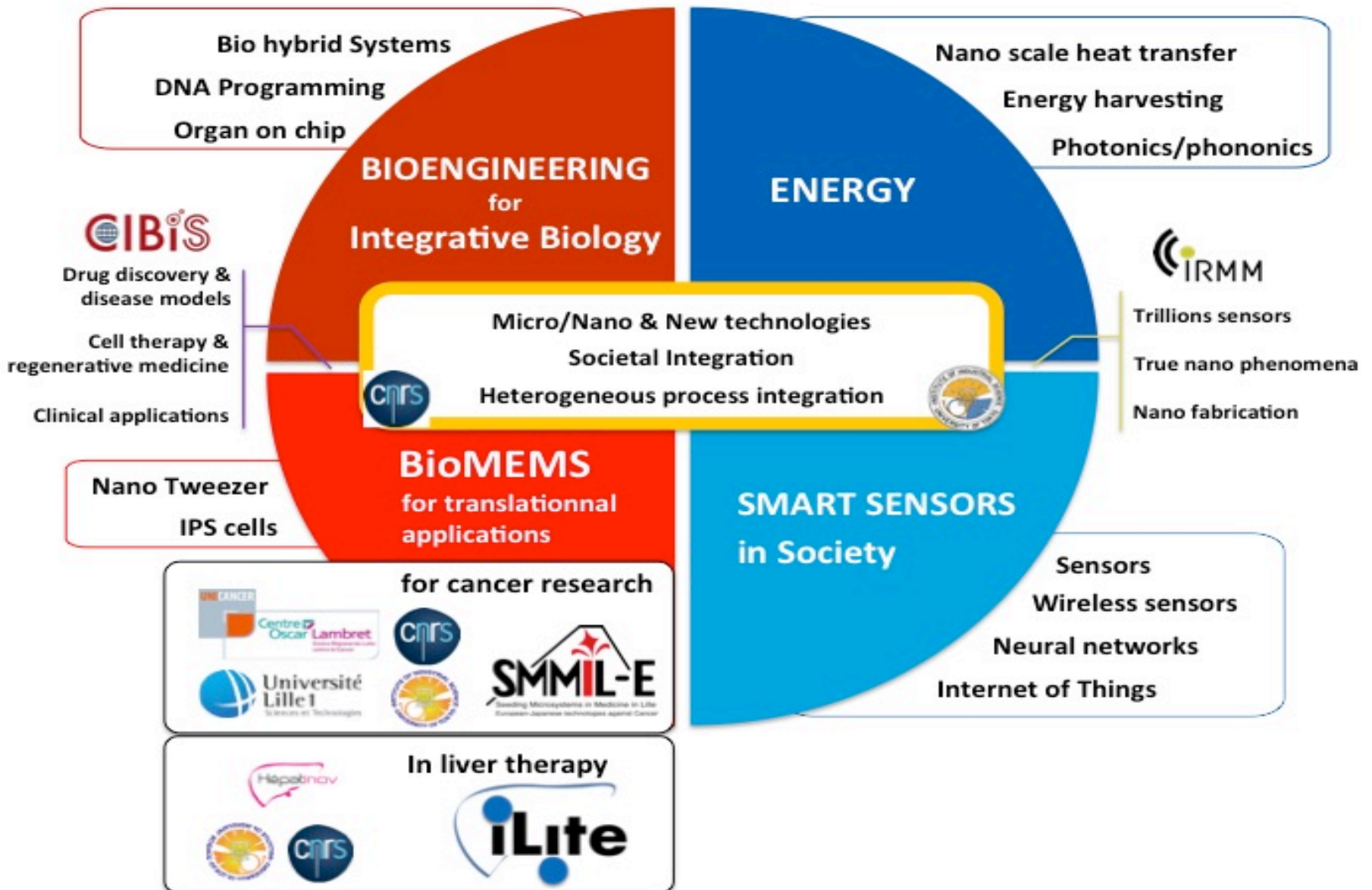


# LIMMS Operation

- Established in **1995**, UMI since 2004, **16 host Japanese laboratories**
- 219** (176 French, 9 Japanese, 26 other nationalities) persons welcomed at LIMMS
  - CNRS senior researchers (37)
  - JSPS Post-Doc fellows (71)** + contract Postdocs (20) + PhD students (13) + Trainees (36)
  - EUJO-LIMMS members (21)**
  - research engineers (5), administrative staffs (15)
  - Industries (2)



# Scientific LIMMS activities



# Translational research - SMMiLE

## Seeding Microsystems in Medicine in Lille – European Japanese Technologies against Cancer

### SMMiL-E:

Mirror structure of LIMMS

↳ IIS research site abroad

- France-Japan collaboration research to implement BioMEMS for cancer therapy
- Located at Centre Oscar Lambret to develop BioMEMS in a hospital
- ONCO-Lille being a major cancer research center in France
- Top ranked hospital in cancer therapy/treatment



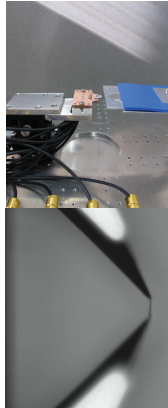
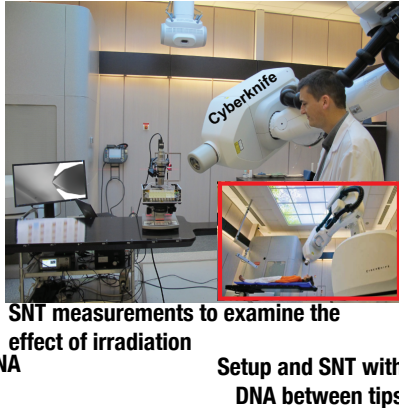
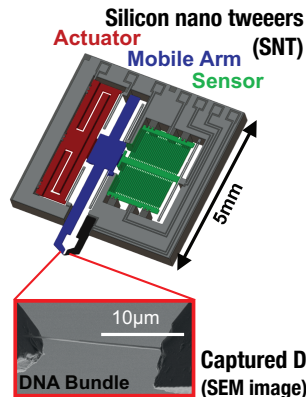


# Translational research - SMMiLE research outputs

## WP1: Resistance in biomolecular mechanism

- DNA degradation under X-ray (radiotherapy)

*Application: Therapy and combined therapies*

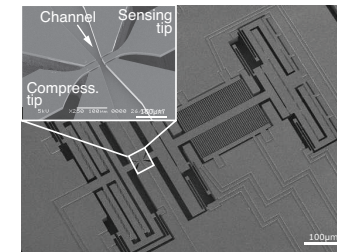
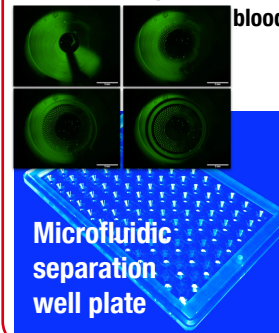


## WP2: Cellular evaluation and diagnosis

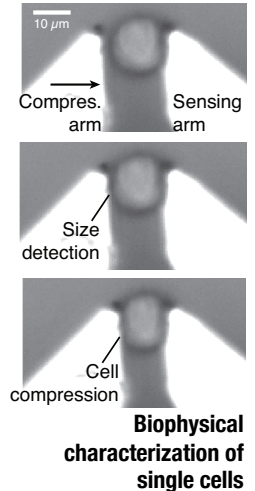
- Circulating tumor cell sorting
- Mechanical characterization of CTCs

*Application: Diagnosis, therapy monitoring*

Cancer cell separation from blood



Cell characterization device with built-in microfluidics



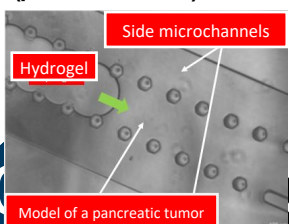
Biophysical characterization of single cells

## WP3: Tumors and therapeutic targets

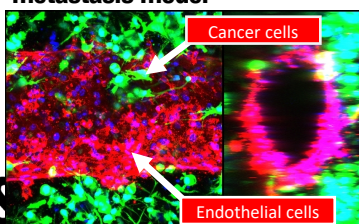
- Developing tumor angiogenesis
- Cellular motility and metastatic processes

*Application: Mechanism of action, drug evaluation*

Organ-on-a-chip (pancreatic cancer)

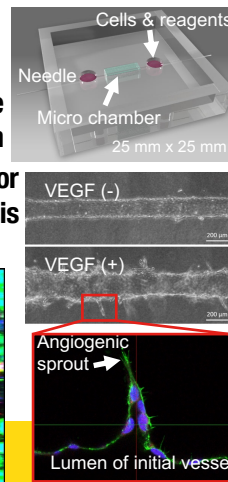


In vitro metastasis model



Vasculature formation

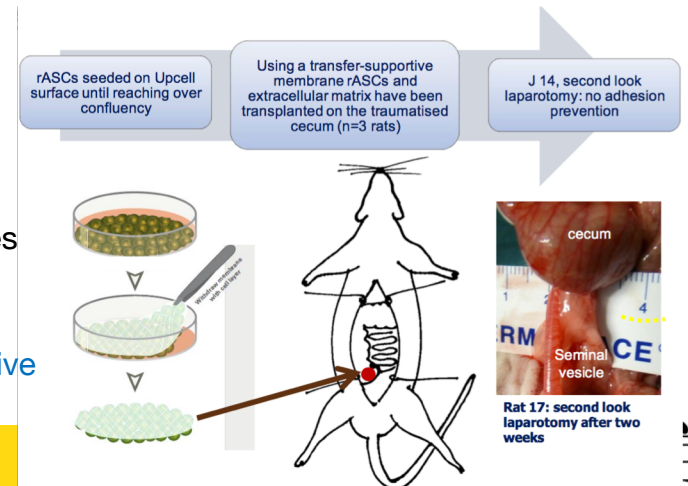
Tumor angiogenesis



## WP4: Biological adhesives and neotissues

- Biological glues
- Cellular fibers
- Patch and new-tissues

*Application: Reconstructive surgery*



# Translational research - SMMiLE main scientific outputs

## Scientific production in numbers:

Research projects (labeled/total): 7/12

Researchers in Lille (current/exp. by oct): 8/14

Research grants (granted/under evaluation): 2/7

Journal articles on related topics: 13

Conf. proceedings (Smmil-e projects): 8

Patents: 2+1

Industrial contact:

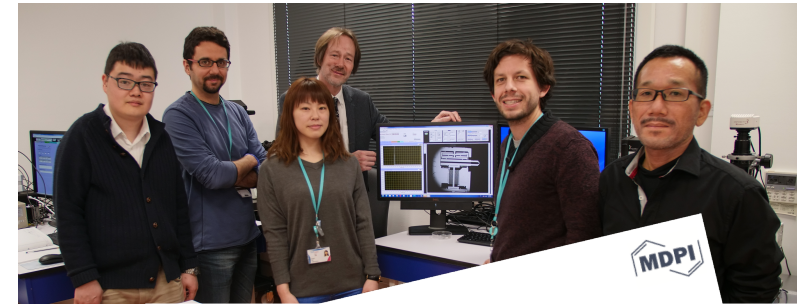
- Start-up company (incubation): 1

- Labcom: 1

Other funding:

- CPER budget: 6M€

- Additional CPER budget:  
(for Recruitment of a young PI) 0.6M€



**Sencet**



# Translational research - iLite



Innovation in Liver Tissue Engineering

- iLite = Innovation in Liver Tissue Engineering technologies for **CLINICAL demand** on liver therapy

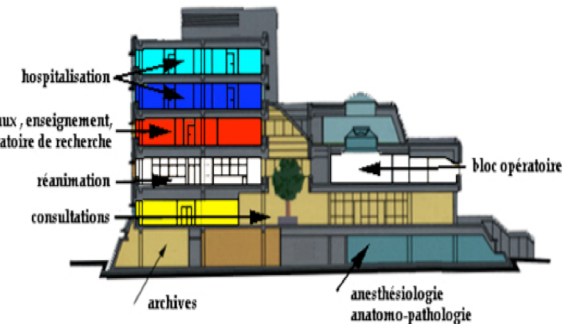
- Biologists, clinicians, engineers
- Public/private consortium (4SMEs)
- 8.5 M euros, 0.4M at LIMMS Tokyo

French and Japanese joint

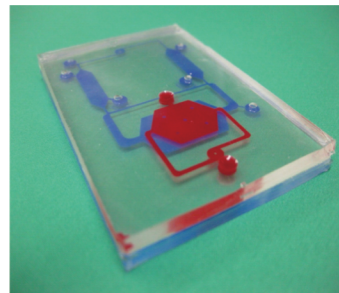
French hospital



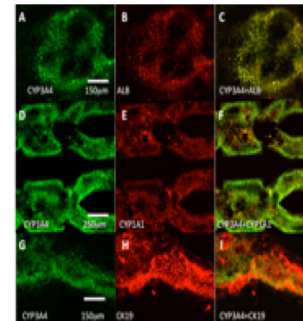
IIS engineering



Clinical Research Facilities



Device Technologies



Organ on a chip,  
Liver iPS  
&  
Innovative  
disease models





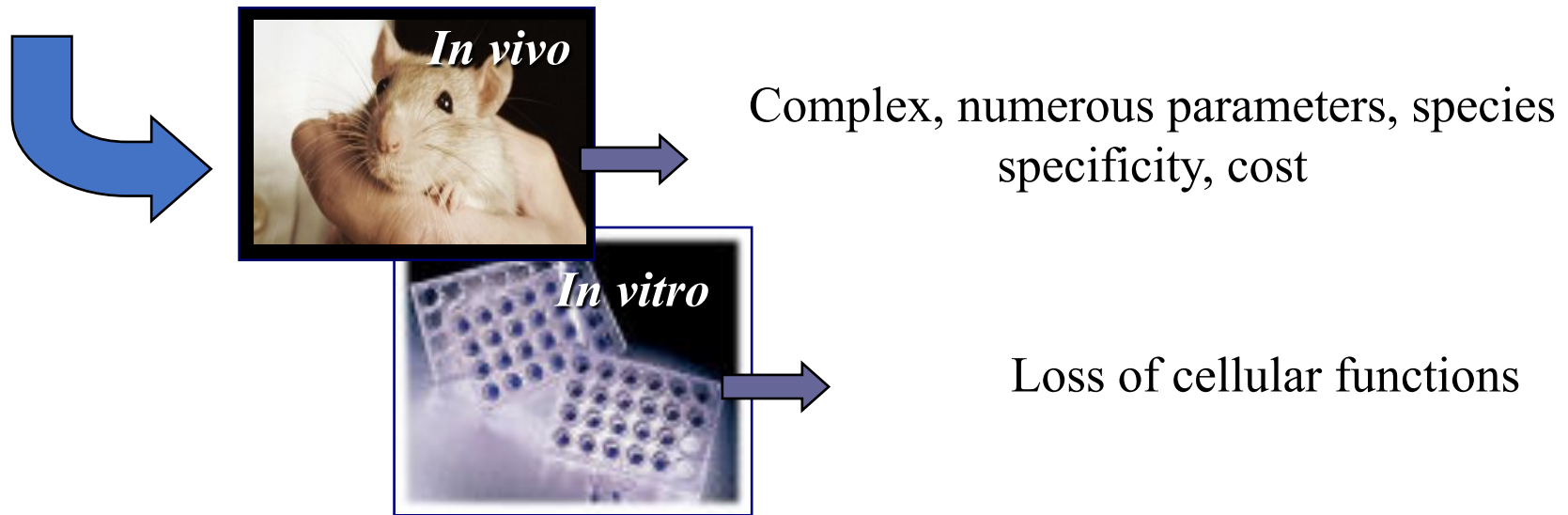
# Translational research - iLite main scientific outputs

- MoU signed : 24<sup>th</sup> of October 2016/ Consortium agreement: October 2017
- 2 LIMMS-IIS publications accepted, 4 submitted, 2 in preparations
- Visit of Pr Legallais, Pr Duclos Vallee at IIS in Dec 2016 /June 2017
- Visit of Pr Sakai, Pr Okitsu at Hepatinov in Feb/Nov 2017
- 3 joint workshops (2 Japan and one in France, 9 Japanese Professors)
- 2 Joint PhD (S Matsumoto: Fujii/Leclerc, A Essaouiba Sakai/Leclerc/Legallais), 3 Post doc fellows, Students exchanges (2 PhD)
- New joint research proposal (RISE, C2C, ANSES, ANR, Kakenhi, ERC)
- Extended collaboration to Univ of Tokyo (Pr Okitsu, Pr Miyajima, Pr Takeuchi)



## Academic and industrial contexts

### How to evaluate the metabolism and effects of xenobiotics

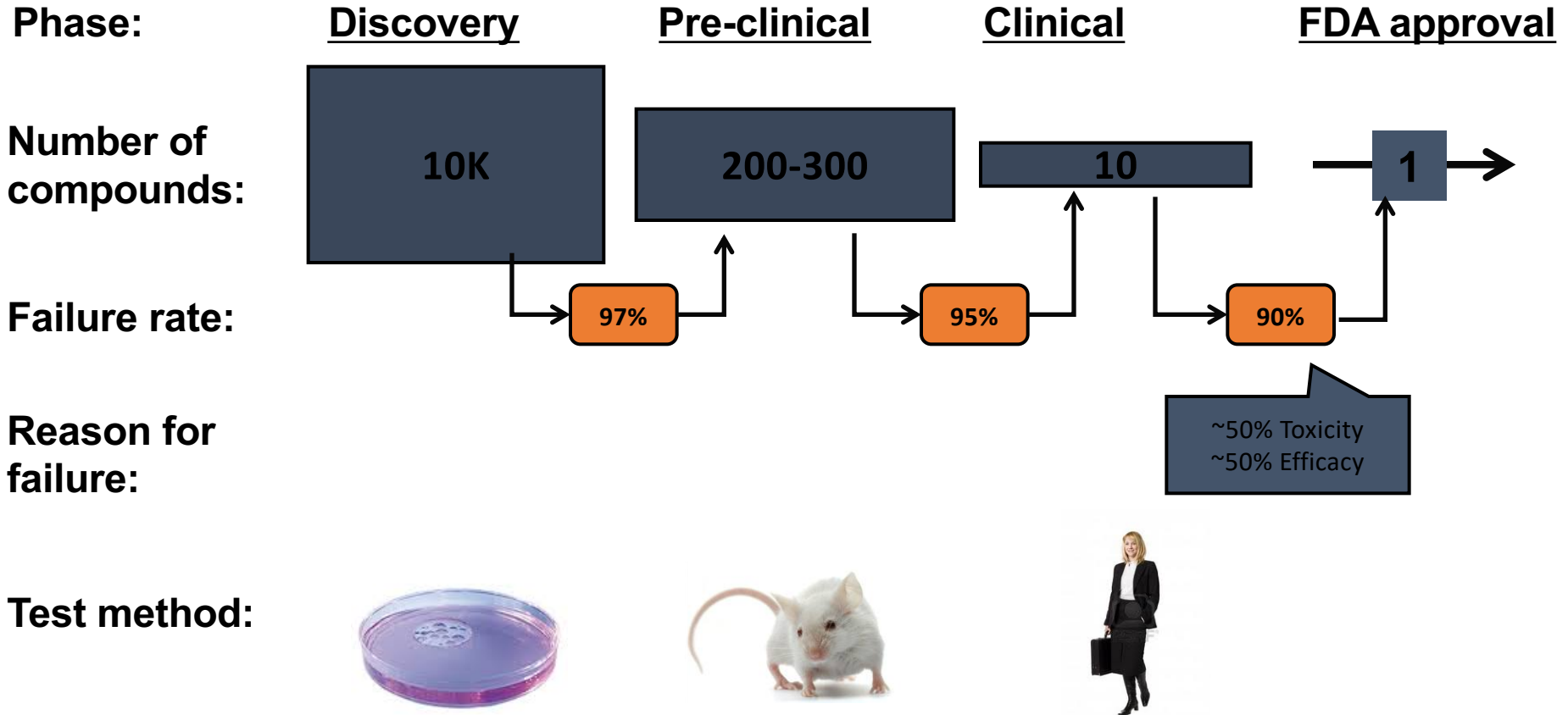


**Industries: cosmetic, pharmaceutical, chemical**

**EU Directive REACH**

# THE PROBLEM

**Time: 10-12 Years**  
**Cost: \$1B**

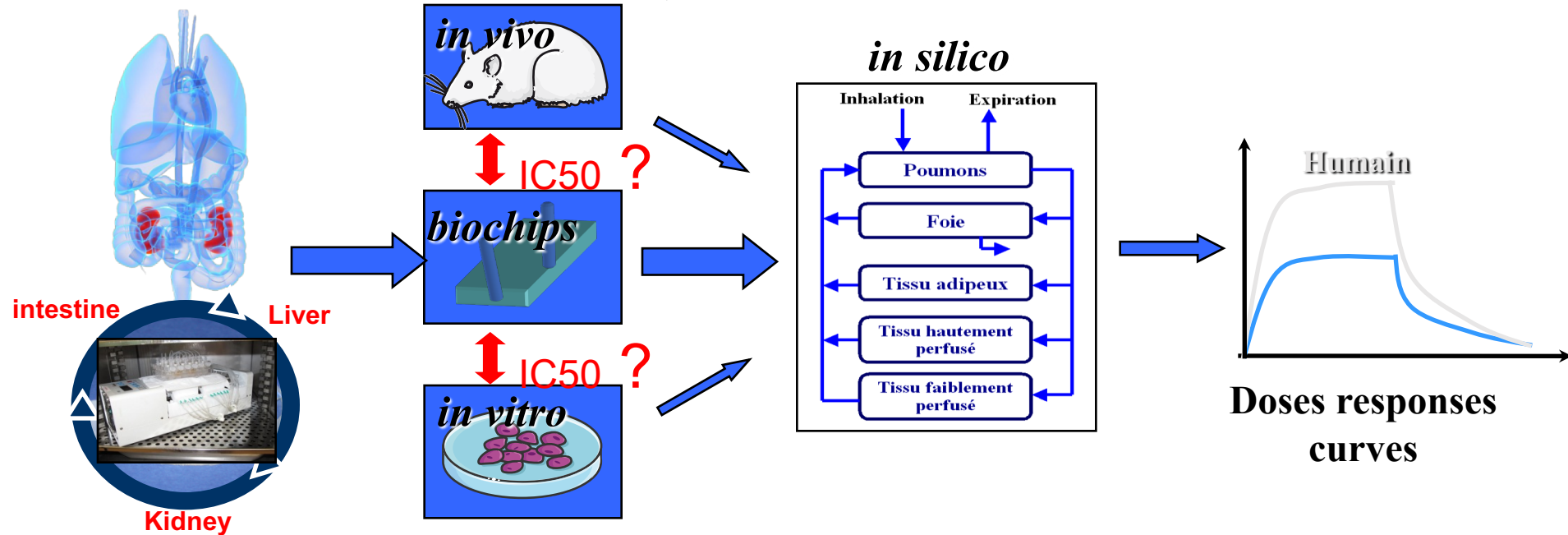




# Human on chip concept

**Objectives:** Better predictivity of toxicity and pharmacokinetics

**Problems :** Pertinence of actual models, costs



## Solutions:

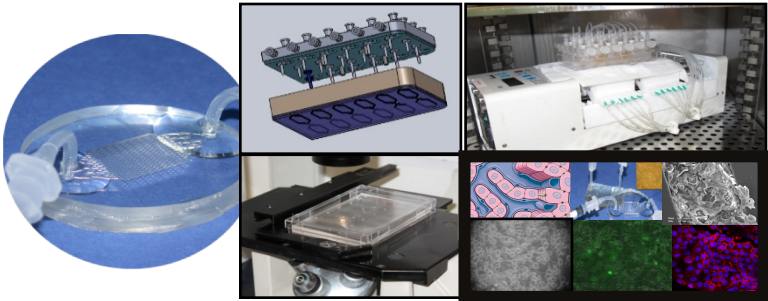
Organ on chip new experimental models; In silico integration

Comparison *in vivo*/ *in vitro* on reference compounds

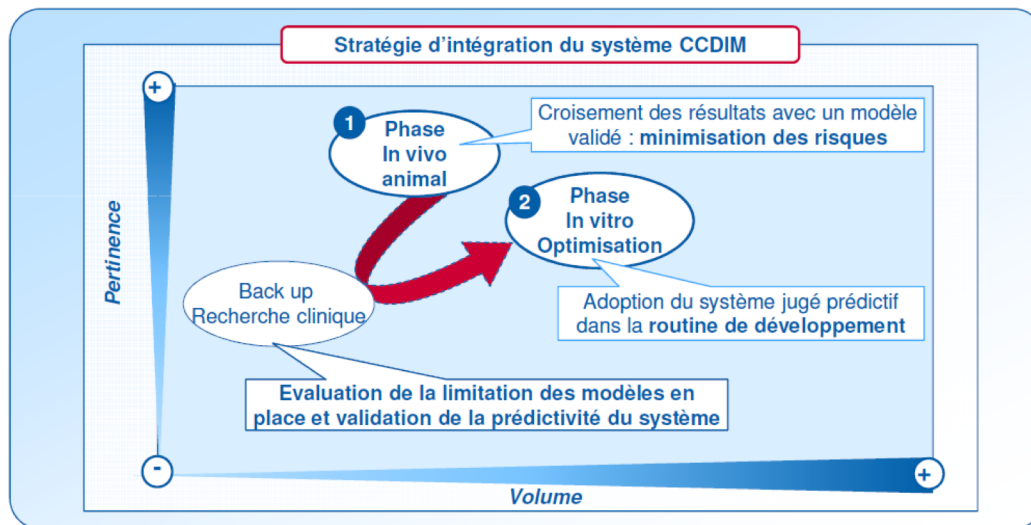
Toward human screening

# Advantages of the microfluidic human like biochips

## Solution : Miniaturisation of the human physiology: human on chip



*miniaturisation* = *parallelisation, cost reduction*  
*design* = *middle throughput (100 molecules after HTP)*  
*integration* = *robotisation, real time analysis*  
*physiology* = *functional cell/tissue*  
*functional* = *Human predictive toxicology*



*in vitro* optimisation  
 =  
 pre clinical back up with marketing values  
 =  
**Costs reduction**  
**False positive reduction**  
**Animal studies reduction**

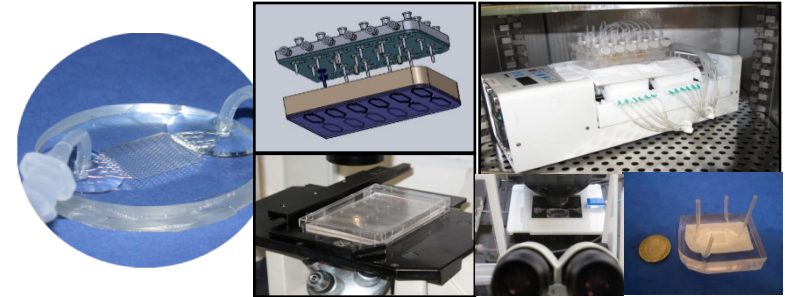
# Overall strategy

## Conception and fabrication of the biochip:

- \* material, design, packaging, models
  - **patent technology (fluidic platform)**

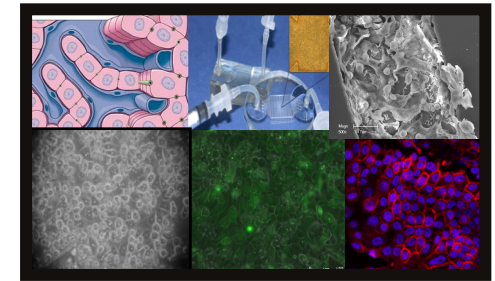
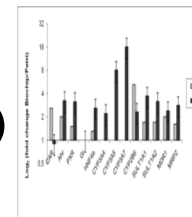
## Bio ingeenering: bioartificial organs:

- \* liver, kidneys, intestine, etc...
- \* cultures protocols (flow rates, cells, etc...)
- \* compatible with biological tools (PCR, etc...)
  - **patent protocol (stem cells)**



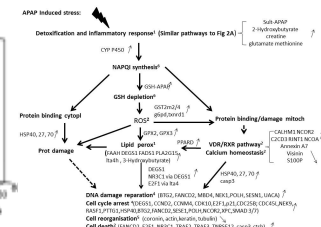
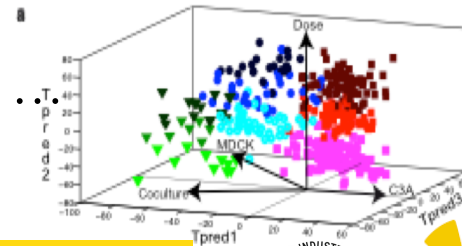
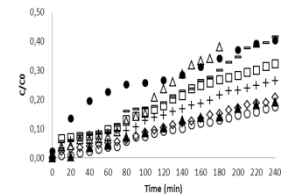
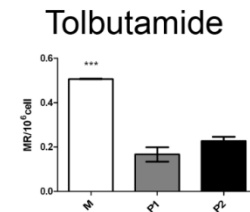
## Functional tissues and biochips

- \* Comparison of biochips, animal and petri data



## Predictive toxicology:

- \* models, omics, biomarkers, gene networks, ...
  - **patent oriented application (screenings)**
  - **know how transfert**





# NEW MOLDED IDCCM CASE AND ITS BIOCHIPS

## Assembling steps of the IDCCM case and the biochips

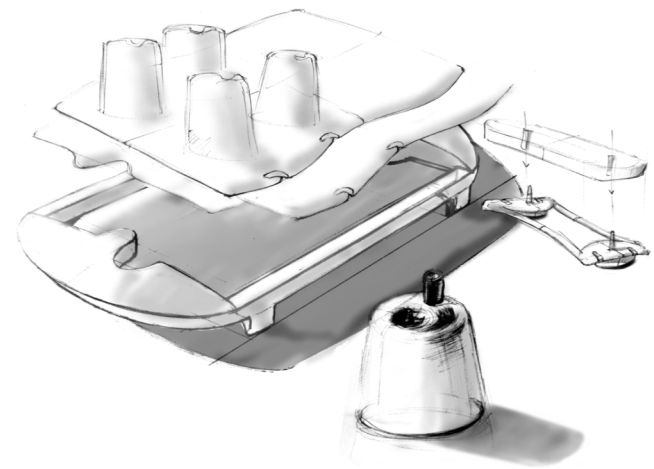
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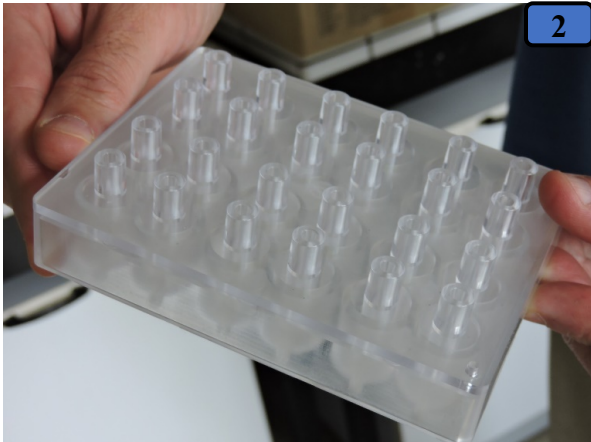
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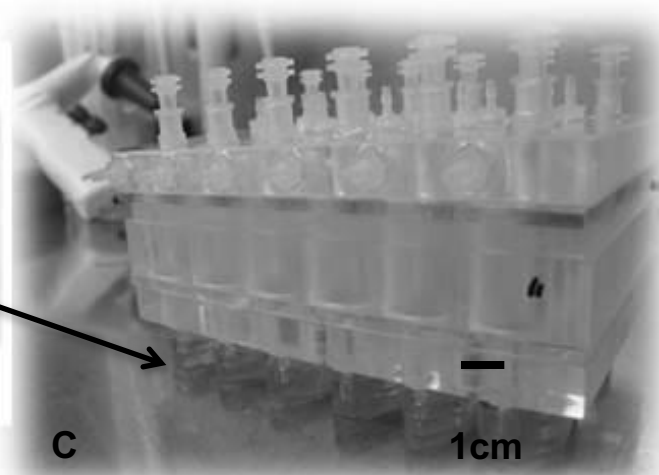
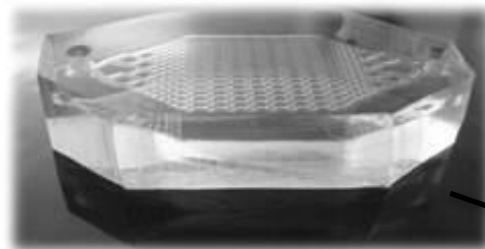
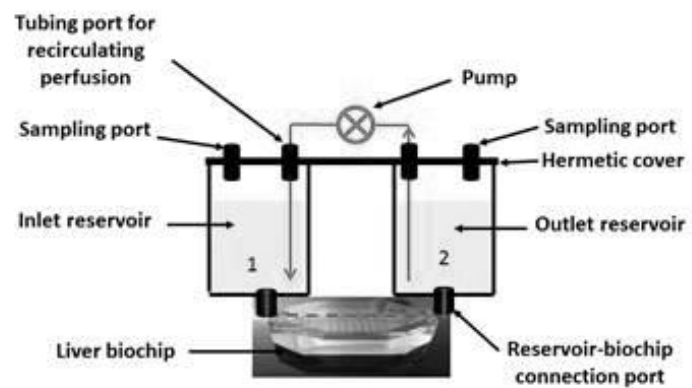


4



2

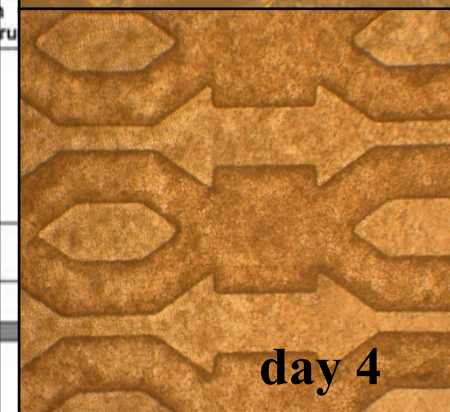
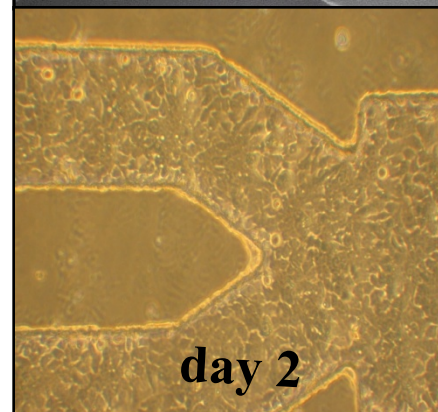
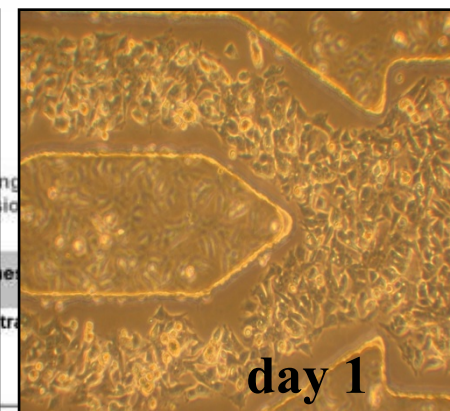
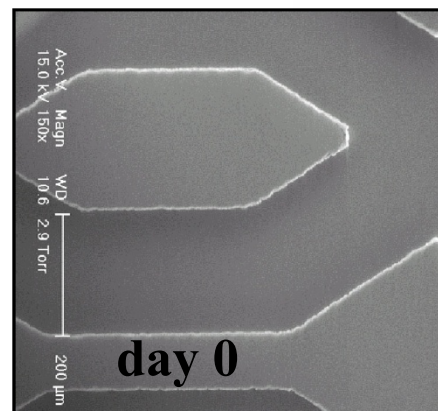
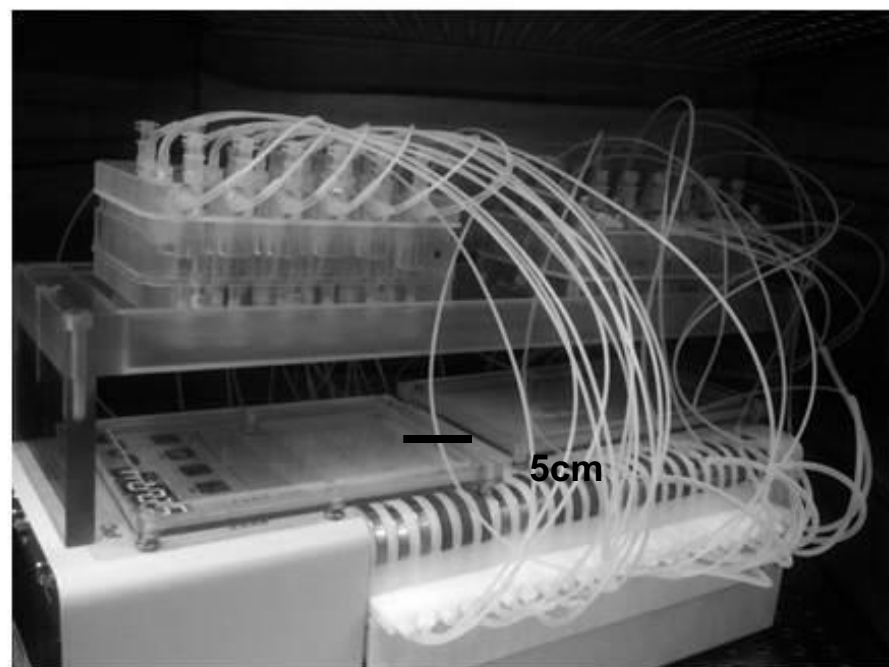




**A**

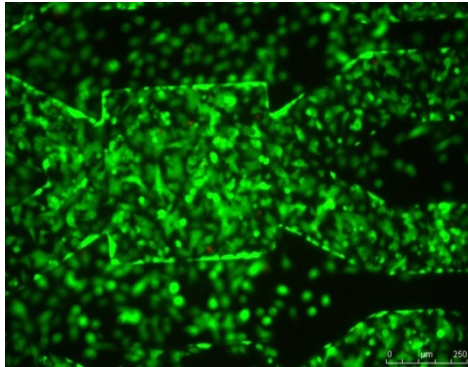
**B**

**C**

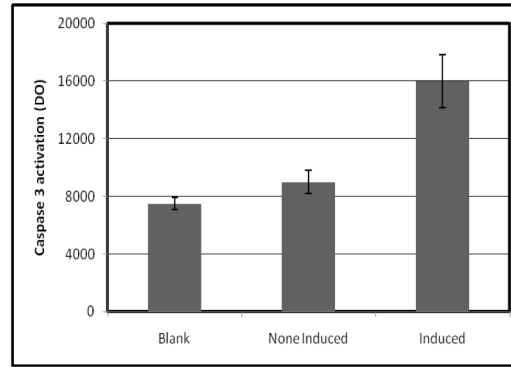


**D** Baudoin et al., JBE 2011; sensor actuator, 2012, biotech prog 2014

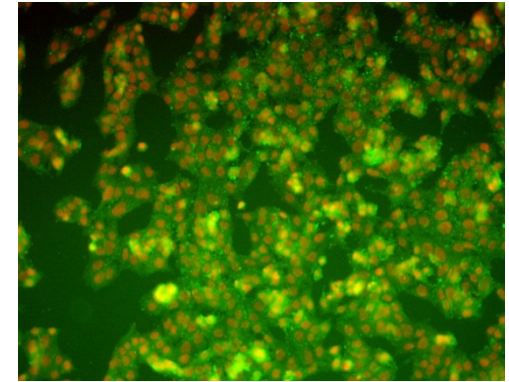
# Viability, basal apoptotic situation in biochips of liver cells



**Calcein AM/IP: viable cells**



**No caspase 3 activation**

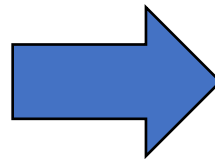


**IP/Annexin V ; ++control (actinomycin D)**

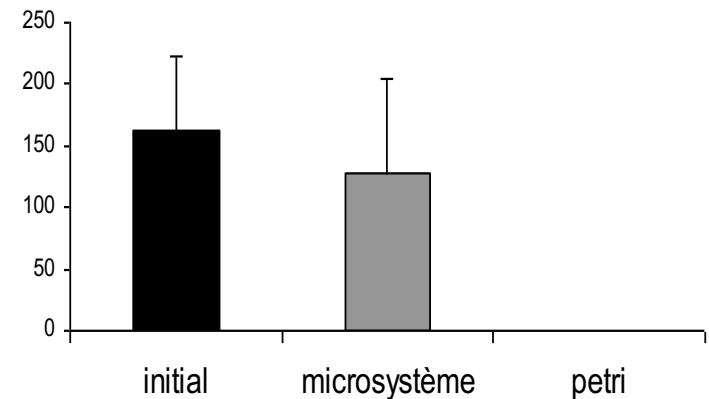
## RTqPCR (U991)

Day 4

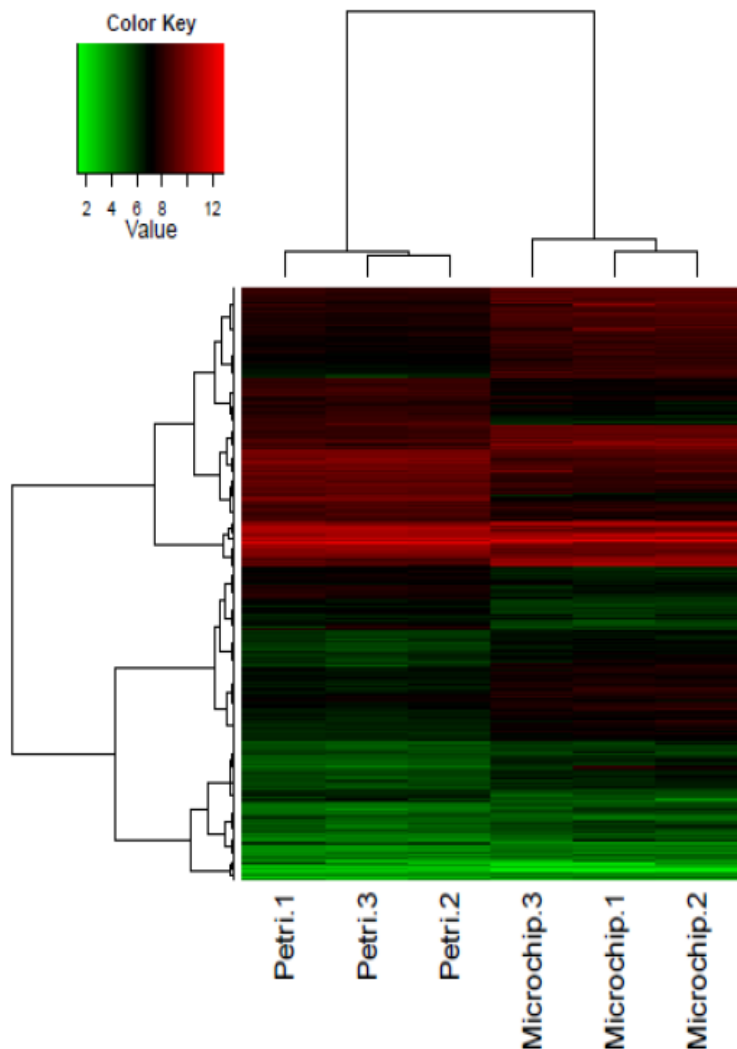
	Petri	Microfluidic Biochip
CYP3A4	100	1078
CYP1A2	100	18011
CYP1A1	100	16903
CYP2B6	100	182
CYP3A7	100	1660
CYP3A5	100	765
SULT1A1	100	353
SULT1A2	100	379
UGT1A	100	791
GR	100	106
CAR	100	71
PXR	100	286
AhR	100	258
HNF4α	100	261
MDR1	100	249
MRP2	100	393



## Activity of CYP1A



# Omics comparison in HepG2/C3a biochips and Petri



At the gene level:

**4012 genes affected** by the microfluidic cultures

At the protein level:

**111 identified proteins** affected by the microfluidic cultures

At the metabolome level:

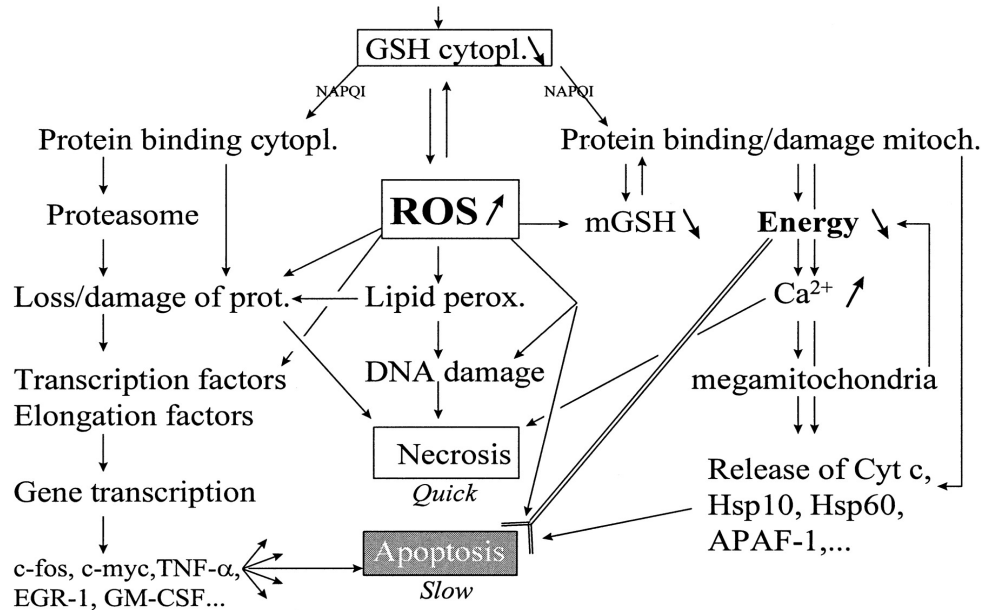
**40 identified metabolites** affected by the microfluidic cultures



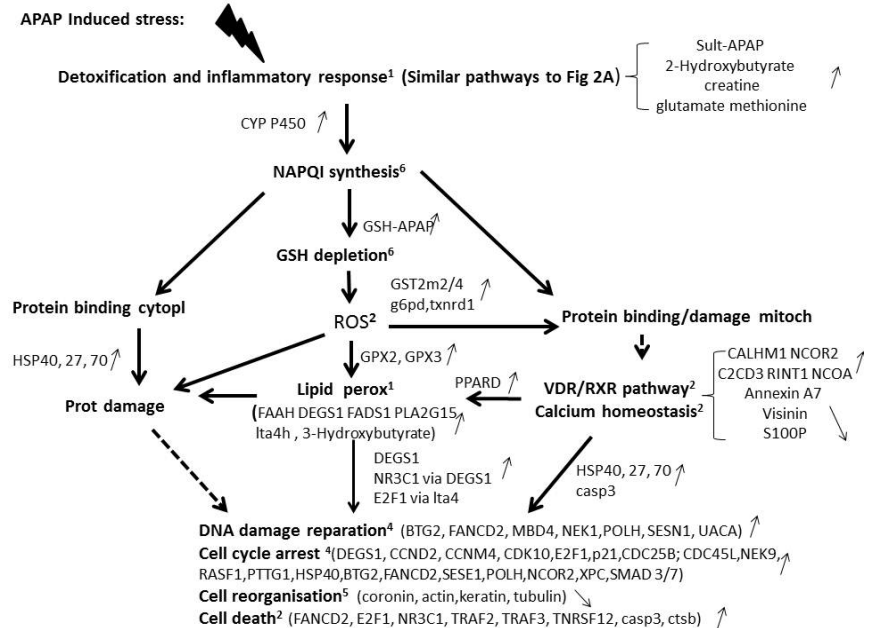
# Pathway and toxic network reconstruction, APAP application

## Context: identification of the mechanism of toxicity

APAP → P450 → NAPQI



APAP *in vivo* data

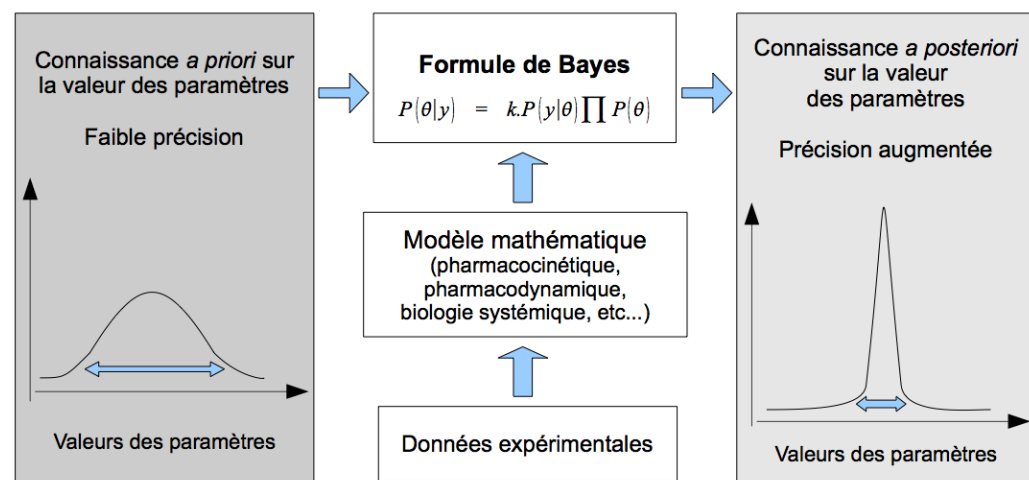
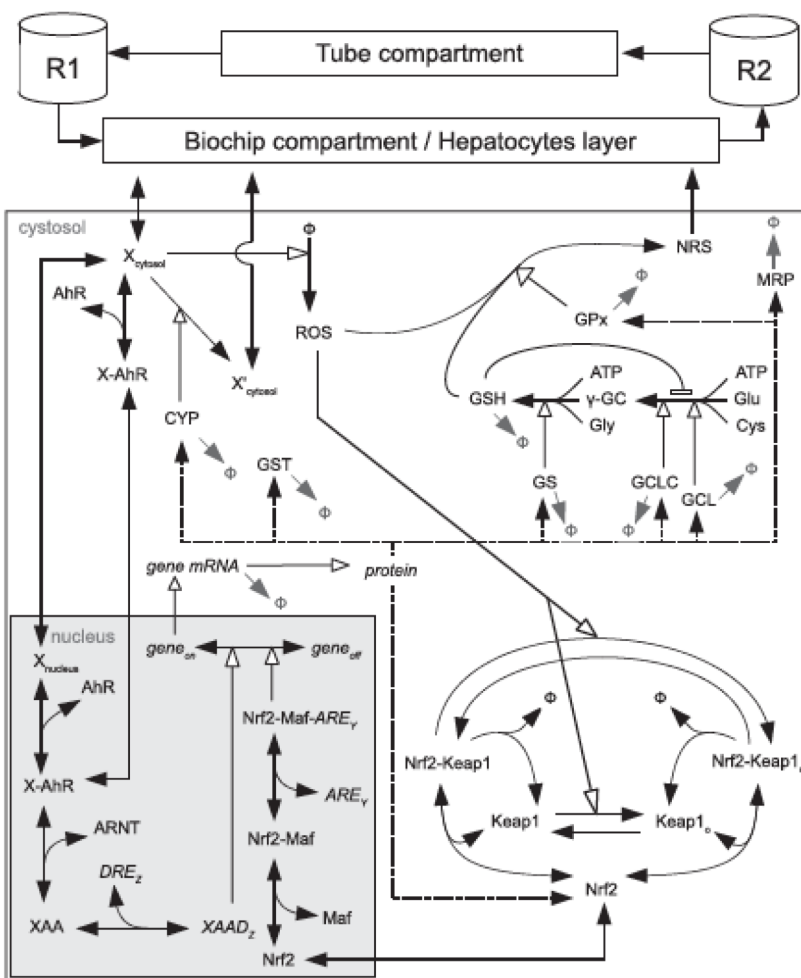


APAP biochip reconstruction

Prot et al, PlosOne 2012; Prot et al., TAAP 2012

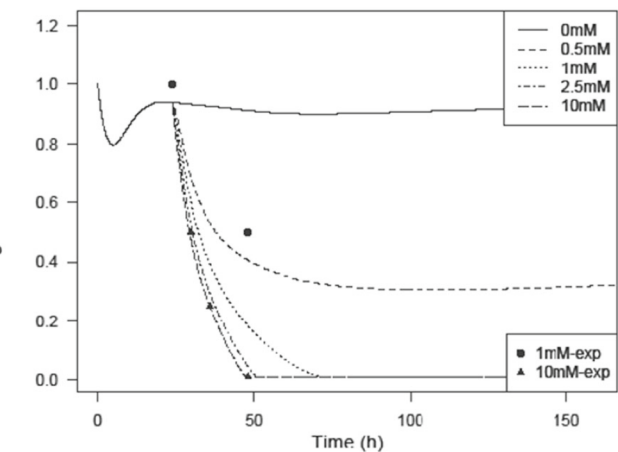
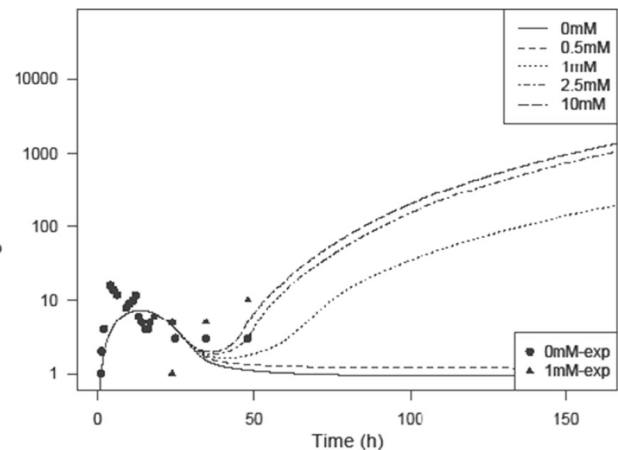
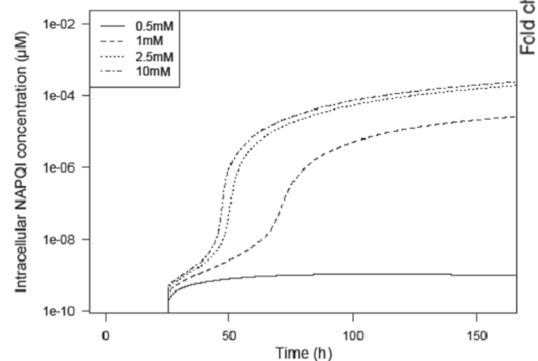
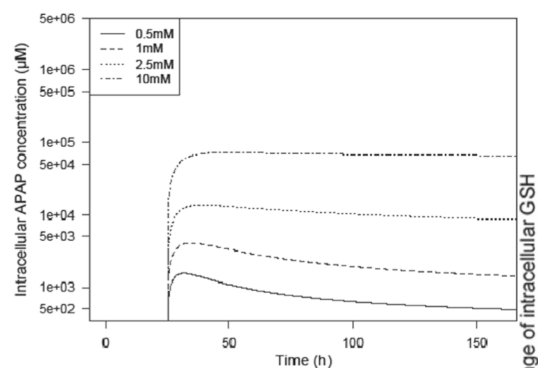
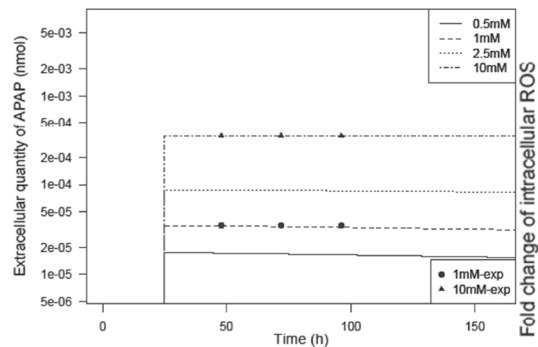
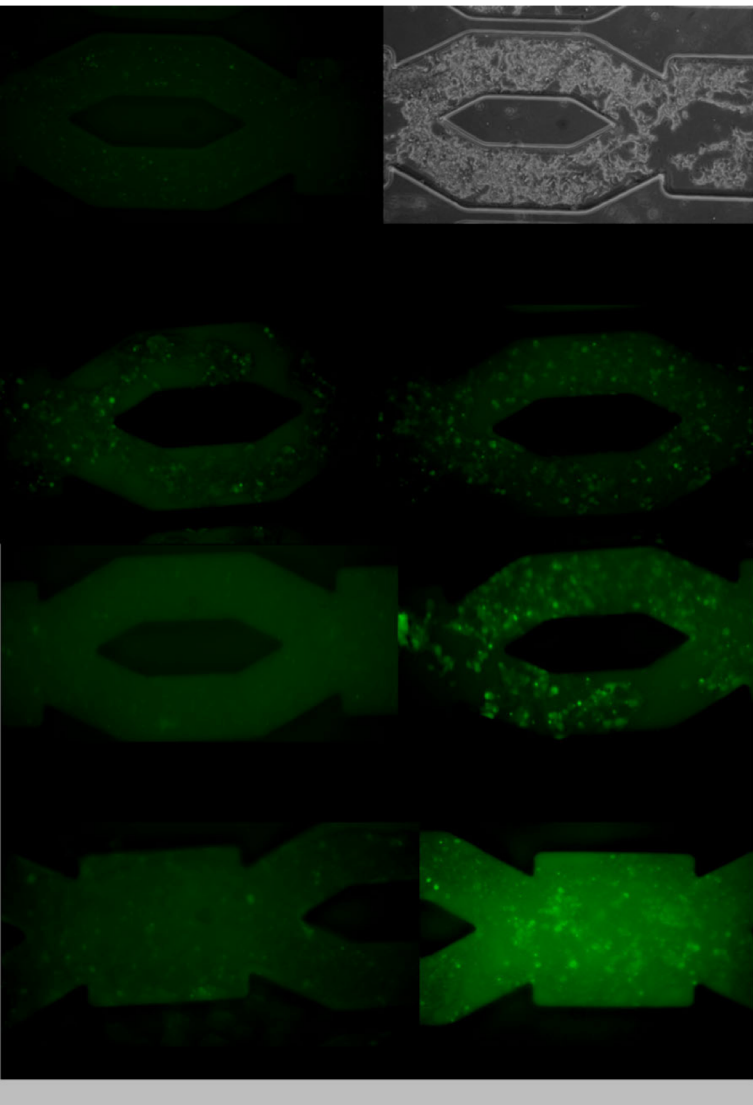


# Modeling strategy based on Bayesian statistics of a PK-Nrf2-GSH model



Leclerc et al; tox in vitro , 2014  
 Leclerc et al. Cell bio tox, 2015  
 Leclerc et al., J applied tox, in press

# APAP-NAPQI toxicity via in silico PK-Nrf2-GSH model in HepG2/C3a

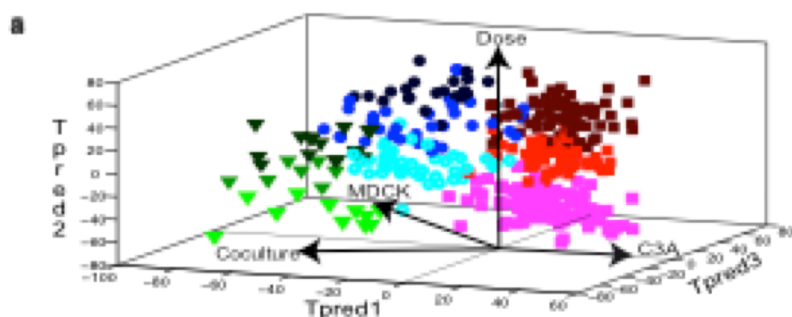


Leclerc et al; tox in vivo , 2014  
 Leclerc et al. Cell bio tox, 2015  
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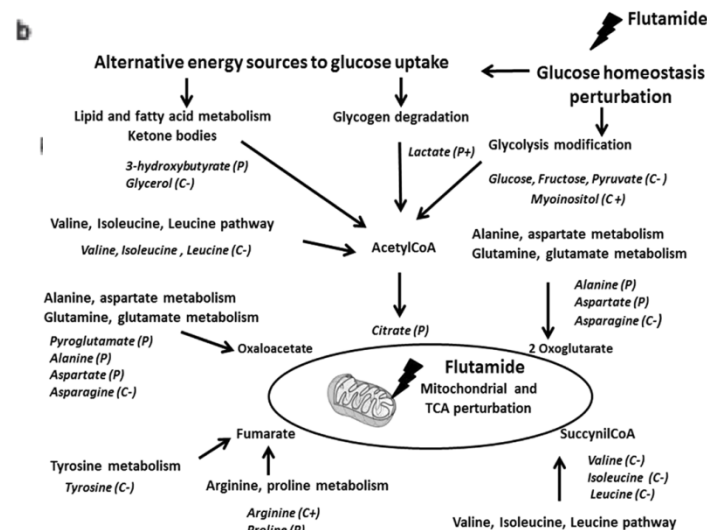
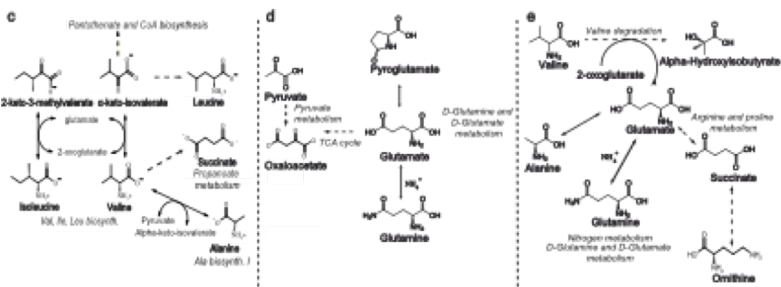
# Metabonomic dose response analysis of chemical toxicity

## Microfluidic signature database:

Liver, kidney, liver – kidney cocultures, Cell lines, primary cells, Petri , biochips  
Molecules : APAP, Flutamine, Hydroxy flutamine, DMSO, NH<sub>3</sub>, MeOH, mixtures



**Dose-trajectory model: dose & compound OPLSDA model**



## Mechanistic interpretation

Coherence with in vivo reports ????

- \*Glucose homeostasis perturbation
- \*Mitochondrial damage

## Specific Molecules signatures

Shintu et al, anal chem 2012; Choucha et al., Tox Sci. 2013

Collab CRMN Lyon



東京大学  
THE UNIVERSITY OF TOKYO



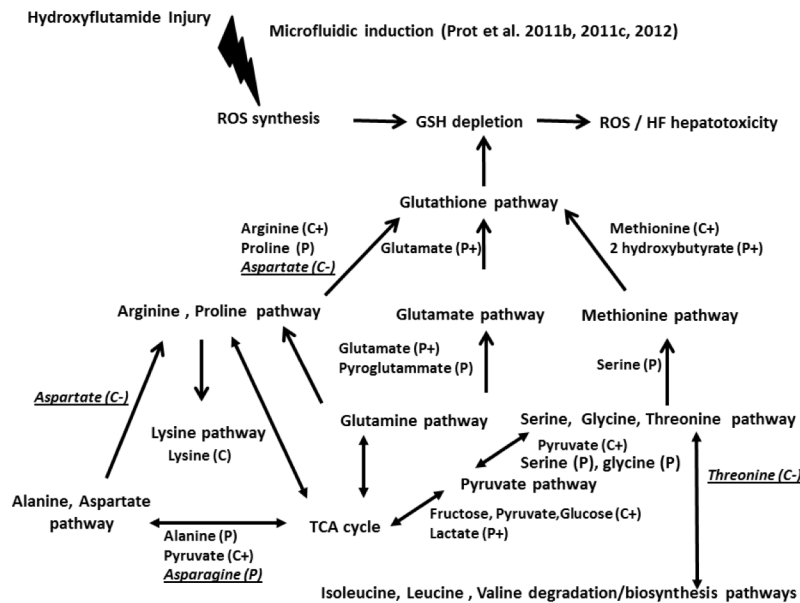
LIMMS  
Laboratory for Integrated Molecular Medicine in the Life Sciences  
UMI 2800



Shintu et al., Anal Chem 2012

# Biomarkers of GSH depletion in HepG2/C3a

## Hydroxyflutamide related GSH Depletion



## Hydroxyflutamide /APAP common biomarkers of GSH Depletion

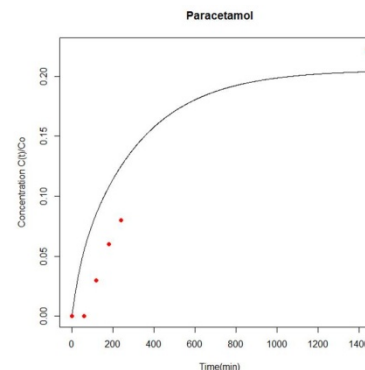
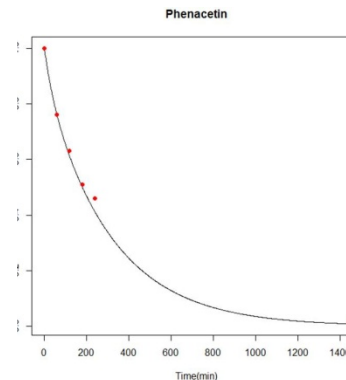
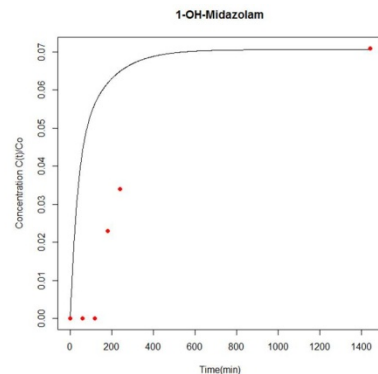
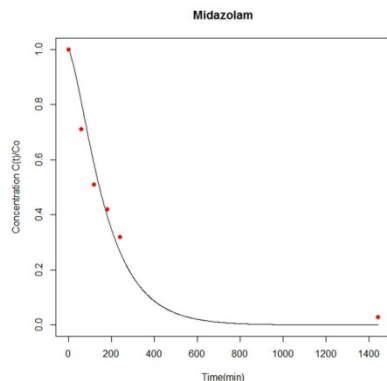
Compounds	Variations	$\alpha_s$ (9 control vs 6 HF biochips)	$\alpha_s$ (9 control vs 11 APAP biochips)
<b>2-Hydroxybutyrate</b>	<b>P+</b>	$3.61 \times 10^{-2}$	$3 \times 10^{-3}$
<b>Alanine</b>	<b>P+</b>	$3 \times 10^{-1}$	$3 \times 10^{-4}$
<b>Arginine</b>	<b>C+</b>	$7 \times 10^{-3}$	$3 \times 10^{-3}$
<b>Fructose</b>	<b>C+</b>	$2 \times 10^{-3}$	$1 \times 10^{-3}$
<b>Glucose</b>	<b>C+</b>	$2 \times 10^{-3}$	$5.5 \times 10^{-3}$
<b>Glutamate</b>	<b>C+</b>	$4 \times 10^{-2}$	$2 \times 10^{-2}$
<b>Lactate</b>	<b>P+</b>	$5 \times 10^{-2}$	$9 \times 10^{-3}$
<b>Lysine</b>	<b>C/C+</b>	$1 \times 10^{-2}$	$5 \times 10^{-3}$
<b>Methionine</b>	<b>C+</b>	$7 \times 10^{-4}$	$5 \times 10^{-2}$
<b>Proline</b>	<b>P</b>	$5 \times 10^{-2}$	$2 \times 10^{-2}$
<b>Pyroglutamate</b>	<b>P</b>	$4.61 \times 10^{-3}$	$6 \times 10^{-2}$
<b>Pyruvate</b>	<b>C+</b>	$6 \times 10^{-1}$	$6 \times 10^{-4}$
<b>Serine</b>	<b>P</b>	$2 \times 10^{-3}$	$2 \times 10^{-2}$

24h-10 $\mu$ M-hydroxyflutamide (HF) 72h-1mM-acetaminophen (APAP)  
Both conditions lead to hepatotoxicity *via* cell death  
when compared to controls.

Choucha et al., Tox Sci. 2013

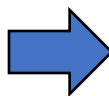


# Drug clearance and extrapolation *in vitro in vivo* in rat



$$\frac{dQ_{HepB}(t)}{dt} = F \times (C_{R1}(t) - C_{HepB}(t)) - fu_{med} \times CL_{int, in vitro} \times C_{HepB}(t)$$

Estimated clearance via *in vitro* PK



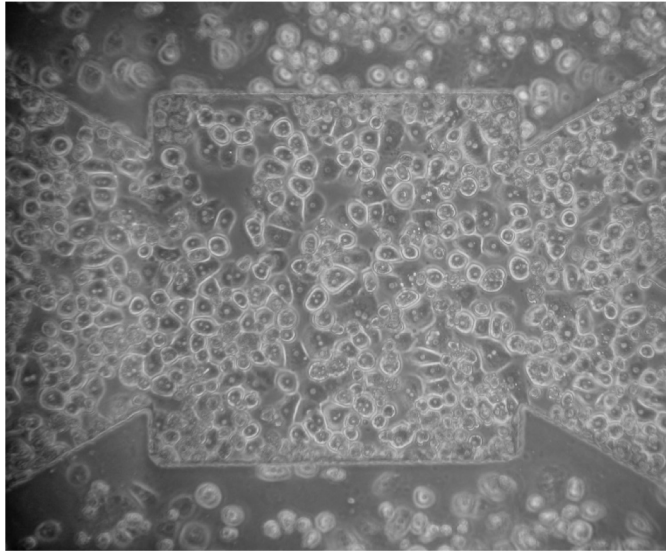
$$CLh = \frac{Q_H \times \frac{fu_b}{fu_{med}} \times CL_{int, in vitro} \times SF}{Q_H + \frac{fu_b}{fu_{med}} \times CL_{int, in vitro} \times SF}$$

Predicted *in vivo* clearance via organ model

	Models predictions (ml/min/kg of BDW)	Literature data (ml/min/kg of BDW)
	<i>CLh in vivo</i>	<i>CLh in vivo</i>
Phenacetin	72	84 <sup>b</sup>
Paracetamol-Phe	41	23.8 <sup>d</sup>
Paracetamol	30	23.8 <sup>d</sup>
Propranolol	85	100 <sup>b</sup>
Tolbutamide	0.1	0.48 <sup>b</sup>

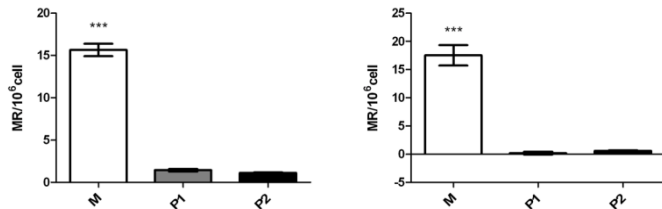
	Models predictions (ml/min/kg of BDW)	Literature data (ml/min/kg of BDW)
	<i>CLh in vivo</i>	<i>CLh in vivo</i>
Caffeine	13.08	12 <sup>b</sup>
Paraxanthine	4.2	15 <sup>a</sup>
Dextromethorphan	61	80 <sup>b</sup>
Dextrorphan	9.5	---
Midazolam	44.1	44 <sup>b</sup>
1 OH-Midazolam	21	62.2 <sup>i</sup>

# Higher metabolic performance in biochips versus Petri for human primary cryopreserved hepatocytes



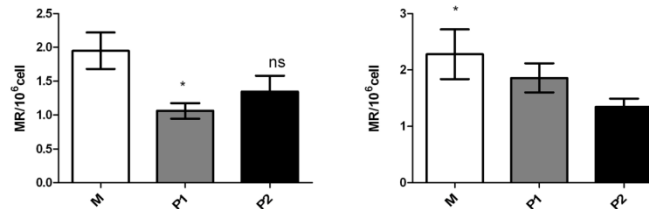
Gene name	post decongelation n=4	Dynamic biochips n=7	Static biochips n=3
HNF4a	100	85±2	6±1
PXR	100	54±1	5±1
CYP1A2	100	80±3	6±1
CYP2B6	100	27±1	1.5±0.2
CYP3A4	100	53±2	0.7±0.2
OATP2	100	98±2	3±0.5
Pgp	100	74±4	74±8

## Dextromethorphan



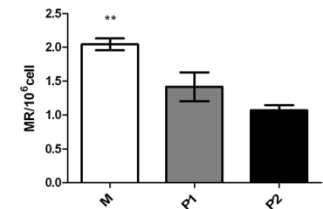
Transformation in Dextromethorphan/  
3-Methoxymorphinan

## Omeprazole



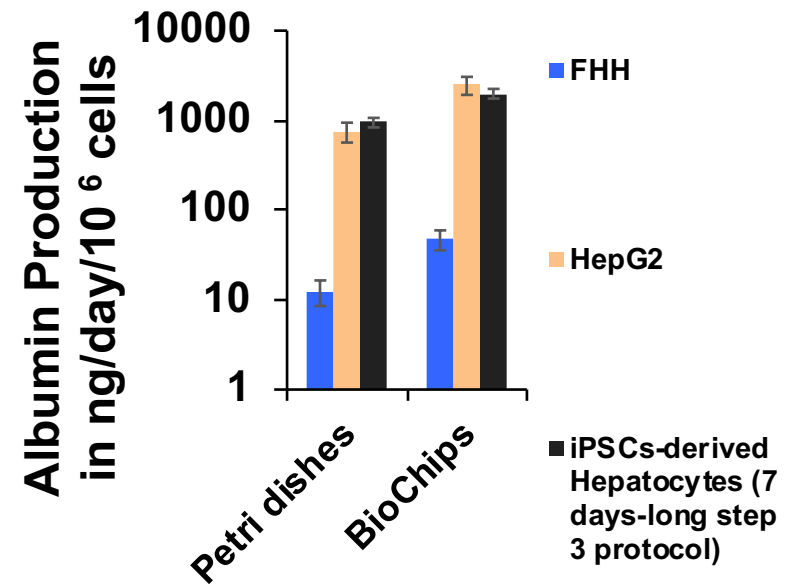
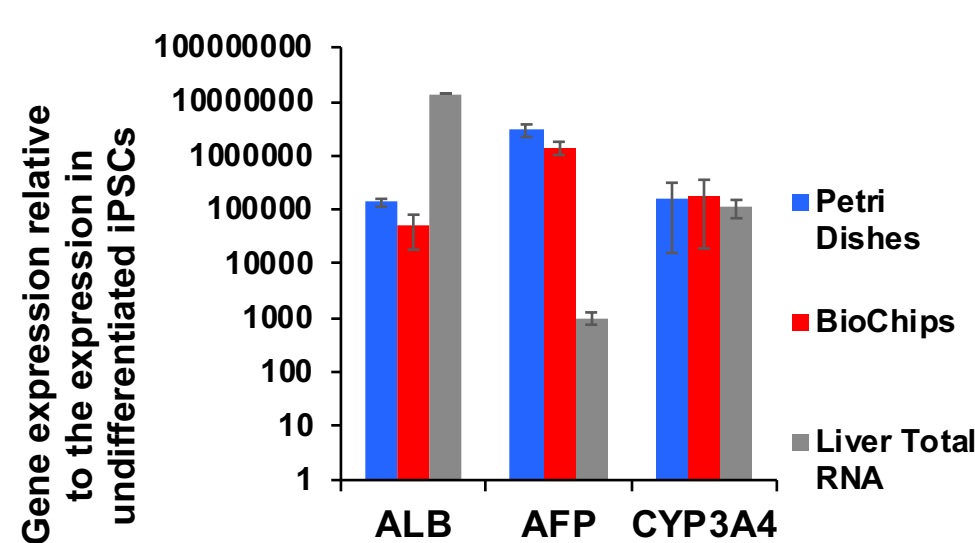
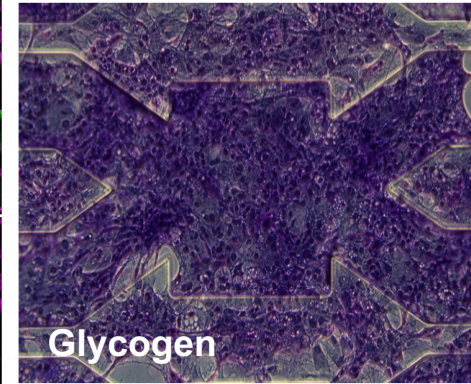
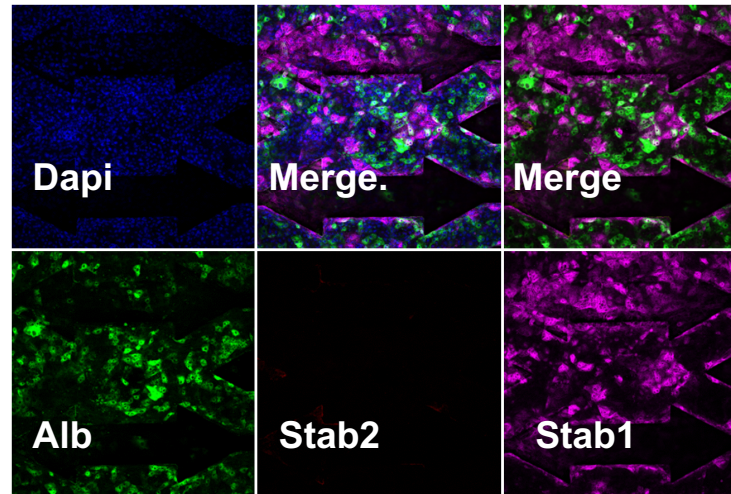
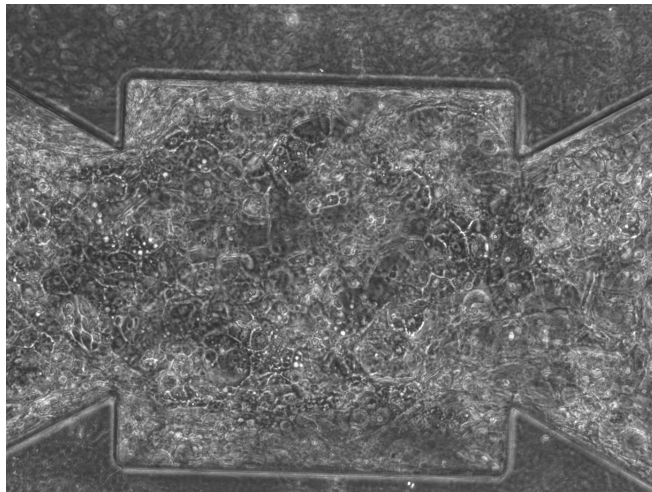
Transformation in 5-OH-  
omeprazole/  
Omeprazole-sulfone

## Acétaminophène



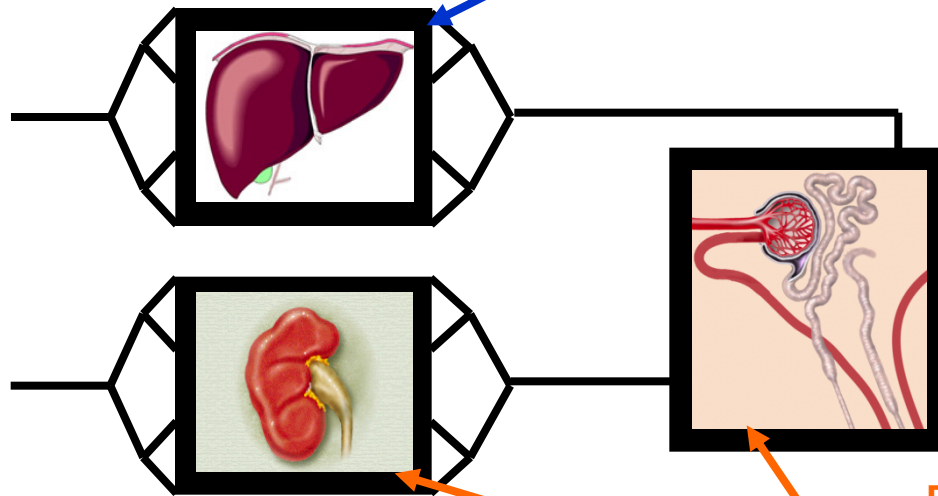
Transformation in  
Acetaminophen-  
glucuronide

# Toward personalised medicine using iPSC



# Liver kidney organs interactions

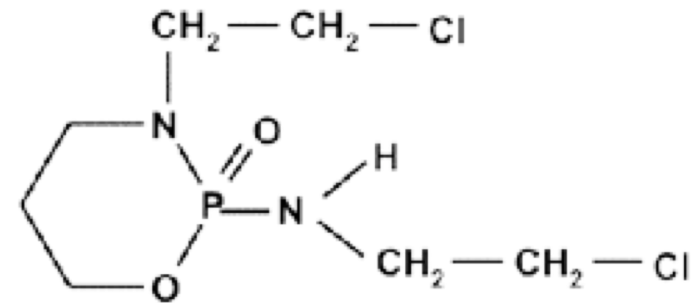
hepatic compartment



Reabsorption unit

Filtration unit

Renal compartments



Ifosfamide

Antinéoplastic drug → Solid tumors

**Nitrogen mustard:**

Alkylation: crosslink with DNA

Apoptosis of cancerous cells

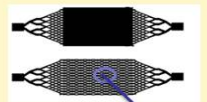
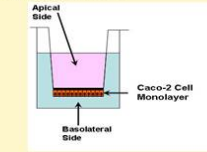
ANR PCV2007  $\mu$ HepaReTox :

UMR CNRS 7338, INERIS (METO), INSERM (991), CNRS UMR 8029 (SATIE BioMIS)



# Integration of barrier in organ on chip module

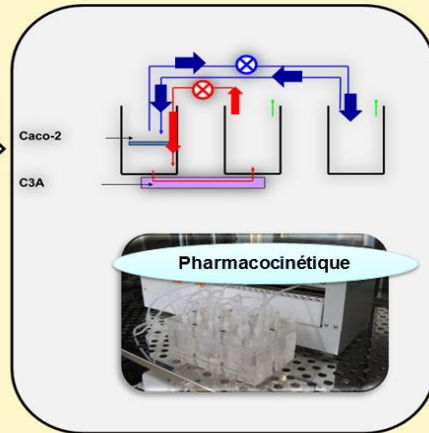
Barrière intestinale  
Lignées CaCO2



Métabolisme  
hépatique Lignées  
HepG2/C3A, primaires

## Couplage intestin – foie

APAP  
Perméthrine  
Effet barrière:  
CTRL +  
CTRL -



8

Barrière  
sertolienne

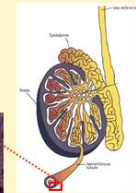
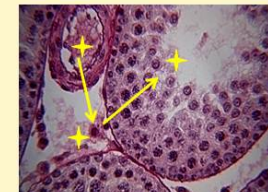
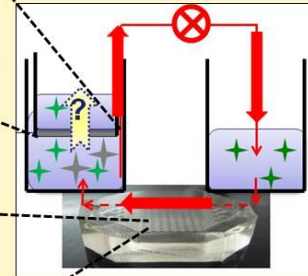
## Couplage foie - testicule

Cellules Primaires

Post-Doctorat 2011 -  
2013



Hépatocytes



## Under going organ to organ interactions:

- intestine liver = first pass
- liver testis = site of toxicity

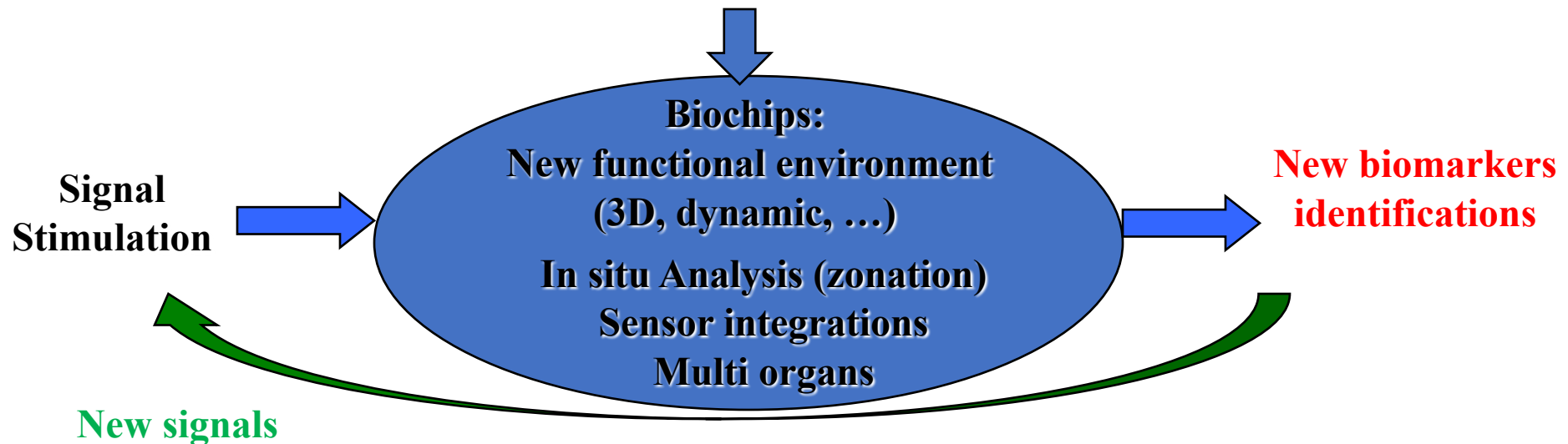
Project Fondation/UTC  
pesticides, drugs

# Microfluidic integrative approach

**Objectives:** Understanding of biological systems

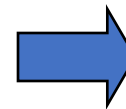
**Problems :** Pertinence of actual model

Conventional and new type of bio analysis



**Solutions:**

On chip system biology  
New signals= new responses



Identification of  
biological processes

# Remerciements

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# Merci de votre attention



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